

Digestion Simulation for Nanoparticle Toxicity Testing: Particle Characterization and *In-vivo In-vitro* Comparison

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Background:

Oral exposure to engineered nanomaterials is increasingly likely due to their frequent use in consumer products. We investigate the influence of simulated gastro-intestinal (GI) pH conditions on particle agglomeration, cytotoxicity and DNA damage induction in intestinal epithelial cell lines and validate the results against *in vivo* DNA damage of the colon.

PVP-capped silver (Ag-PVP) and titanium dioxide (TiO₂) nanoparticles (NP) were tested on Caco-2 and HT29-MTX-E12 (E12) cells as well as in 28-day feeding studies. The pesticide chlorpyrifos (CPF) served as a non-particulate control both *in vitro* and *in vivo*.

The results are part of a pilot study to facilitate studying the effects of ENM in advanced *in vitro* models of the intestine.

Conclusions & Outlook

- Simulation of GI pH conditions induced agglomeration of Ag-PVP and TiO₂ NP, but did not influence the average toxicity of the NP
- The simulated digestion reduced the cytotoxic potential of CPF in Caco-2 cells
- TiO₂ NPs caused moderate DNA damage in Caco-2 and E12 cells in absence of cytotoxicity, and no effects in murine colon cells
- Ag-PVP NP caused pronounced DNA damage with & without concurrent cytotoxicity in Caco-2 and E12 cells, respectively, but no effects in murine colon cells
- Digestion simulation can enhance the applicability of *in vitro* methods and will be adapted for use in an advanced intestinal co-culture model (Fig. 1) within the PATROLS project*

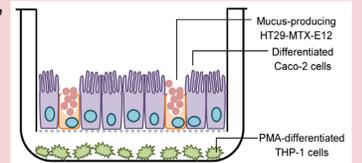


Fig. 1 Advanced intestinal co-culture

Simulation of gastro-intestinal pH conditions

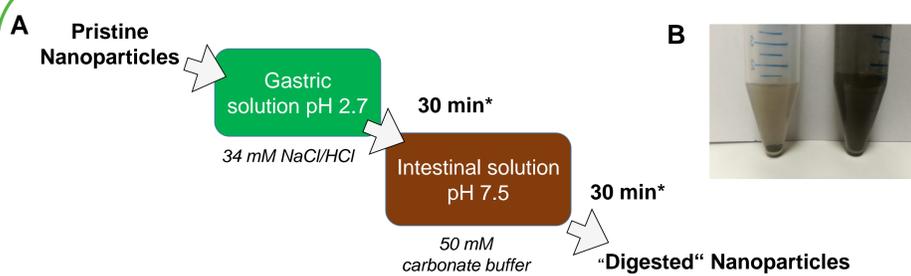


Fig. 2 (A) Schematic description of the artificial digestion process mimicking the gastro-intestinal pH conditions (Method based on: Gerloff et al. 2013 Nanotoxicology); (B) Example of Ag-PVP NP before (right) and after (left) digestion [*Shaking with 120 rpm at 37 °C]

Nanoparticle characterisation

Tab. 1 Charakterisation of Ag-PVP and TiO₂ NP

| Nanomaterial | SEM Ø [nm] | Z-average (DLS) [nm] | D50 (DLS) [nm] |
|-----------------------|-------------|----------------------|----------------|
| Ag-PVP (UD) | 888 ± 1181 | 773 | 472 |
| Ag-PVP (D) | 200 ± 137 | 29219 | 5040 |
| TiO ₂ (UD) | 438 ± 586 | 627 | 567 |
| TiO ₂ (D) | 1539 ± 1601 | 2196 | 1823 |

*UD: undigested, D: digested

- SEM Ø based on analysis of 300 agglomerates
- Simulation of GI-pH conditions caused a strong increase in hydrodynamic diameter in Ag-PVP NP, but only a moderate increase in TiO₂ NP
- SEM analysis confirms size increase for TiO₂ NP, but not Ag-PVP NP

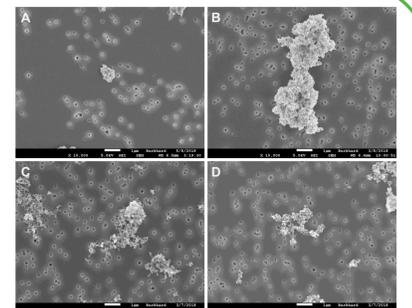


Fig. 3 SEM images of Ag-PVP NP before (A) and after (B) digestion and TiO₂ NP before (C) and after (D) digestion

In vitro: Cytotoxicity

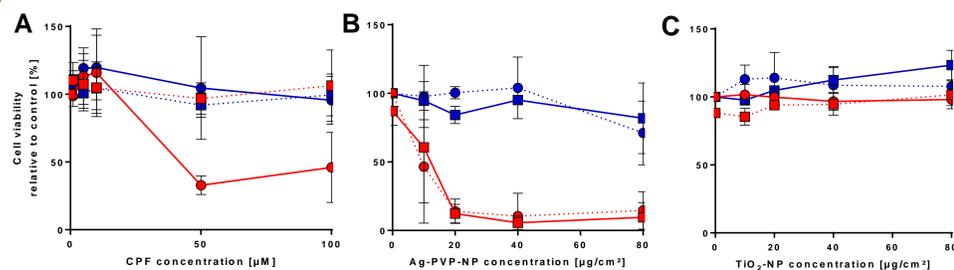


Fig. 4 Metabolic activity in Caco-2 (red) and E12 (blue) cells after 24h exposure to (A) CPF, (B) Ag-PVP NPs, and (C) TiO₂ NPs in their pristine form (circles) or after incubation in simulated gastro-intestinal pH conditions (squares) (Average ± SD of N=3)

In vitro: DNA damage

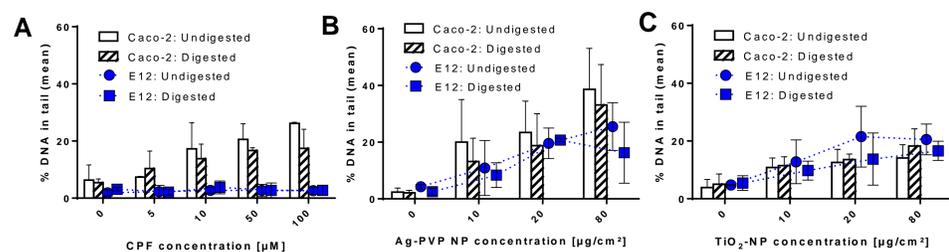


Fig. 5 DNA damage in Caco-2 and E12 cells after 4h exposure to (A) undigested (Caco-2: white bars; E12: blue circles) or digested (Caco-2: striped bars; E12: blue squares) CPF, (B) PVP-capped Ag NPs or (C) TiO₂ NPs. DNA damage was assessed using the alkaline comet assay. (Caco-2: CPF: Average±SD N=2, Ag-PVP & TiO₂: N=3; E12: Average±SD N=2)

In vivo: DNA damage in colon epithelial cells

- 2 cm of proximal colon was cut open, fixated with a microscopic glass slide and epithelial cells were scraped off using as second slide
- The cell suspension was homogenised and pressed through a 70 µm nylon filter
- After centrifugation the supernatant was discarded and the cell pellet re-suspended in 1 ml buffer.* *Method based on: Risom et al. 2007 Toxicology Letters

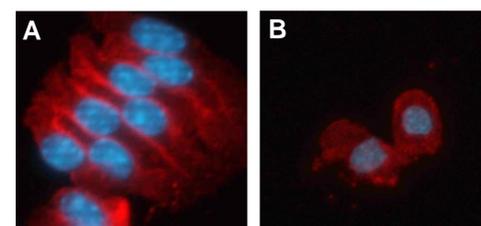


Fig. 6 CD326 Immunostaining of isolated murine colon epithelial cells (40x magnification). (A) intact connective epithelial tissue; (B) single cells as used for Comet Assay (Red (AlexaFluor 594): Epithelial cell adhesion molecule (EpcAM), Blue (Hoechst): nuclei)

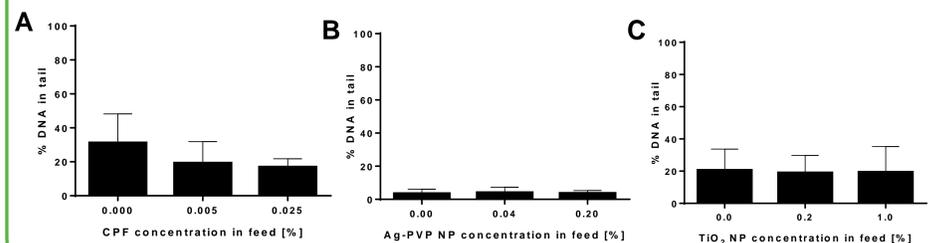


Fig. 7 DNA damage in isolated murine colon epithelial cells after 28-day feeding studies exposing mice to (A) CPF (0,005 or 0,025 % in feed), (B) Ag-PVP NPs (0,04 or 0,2 % in feed) and (C) TiO₂ NPs (0,2 or 1 % in feed) (Average±SD of 5 animals per group; control animals were included in each feeding study)

Materials & Methods

Ag-PVP (Sigma, <100 nm) and TiO₂ NP (P25) were suspended in H₂O (4 mg/mL) and sonicated according to the NanOxiMet suspension protocol (www.nanoximet.eu).

The particle characterisation using Dynamic Light Scattering (DLS) and Scanning Electron Microscopy (SEM) was performed by the Institute of Energy and Environmental

Technology e. V. (IUTA, Duisburg, Germany).

Caco-2 and HT29-MTX-E12 cells were grown for 48 h before treatment. 16 – 20 h before treatment, the cell culture medium was changed to MEM (Caco-2) or DMEM (HT29-MTX-E12) containing 1 % FCS. Cytotoxicity was assessed via **WST-1 assay**, using the Roche Cell Proliferation the Ready-To-Use

Reagent WST-1 after 24 h of treatment.

DNA damage was analyzed via **alkaline comet assay** after 4 h of treatment.

For the **feeding studies**, five week old C57BL/6J mice were used. Five animals per concentration were fed for 28 days before euthanization and dissection.

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* Physiologically Anchored Tools for Realistic nanomaterial hazard assessment

